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NEWS EXPRESS OCTOBER 29 CURRENT WINDOWS VERSION IS V7.01A, CURRENT
MACINTOSH VERSION IS V6.0C(ENG) AND V6.0JC(JP),
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BIOTECCHNO, CAB, CANCERLIT, CAPLUS, CEBA-VTB, CIN, CONFSCI, CROPB,	
CROPU, DDBF, DDFU, DGENE, DISSABS, ...	

2004

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=> s (bacteriophage or phage) (w) 77

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10	FILE BIOSIS
2	FILE BIOTECCHAS
2	FILE BIOTECCHDS
1	FILE BIOTECCHNO
11	FILE CAPLUS

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3	FILE SCISEARCH
2	FILE TOXCENTER
4	FILE USPATFULL
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L1 QUE (BACTERIOPHAGE OR PHAGE) (w) 77

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L4 22 L3 NOT PELLETIER?/AU
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SEARCH ENDED BY USER
=> s l4 and sequence
L5 5 L4 AND SEQUENCE
=> d 15 bib ab 1-5
L5 ANSWER 1 OF 5 CAPLUS COPYRIGHT 2004 ACS ON STM
AN 2003:6130 CAPLUS
DN 138:69476
TI Method for separating bacterial cells and bacterial cell components using
Immobilized bacteriophage or bacteriophage proteins
IN Karelh, Michael Schuetz; Grassl, Renate; Meyer, Roman; Frick, Sibylle;
Robl, Ingrid; Zander, Thomas; Miller, Stefan
PA Profos A.-G., Germany
SO PCT Int. Appl., 40 pp.
CODEN: PIXXD2
DT Patent
LA German
LA German
PATENT NO. KIND DATE APPLICATION NO. DATE
P1 WO 2003000888 A2 20030103 WO 2002-DE2302 20020624
WO 2003000888 A3 20031224
W: AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN,
CO, CR, CU, CZ, DE, DK, DM, DZ, EC, EE, ES, FI, GB, GD, GE, GR,
GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR,
LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MY, NZ, NO, NZ, OM, PH,
PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TN, TR, TT, TZ,
UA, UG, US, UZ, VN, YU, ZA, ZM, ZW, AM, AZ, BY, KG, KZ, MD, RU,
TJ, TM
RW: GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZM, ZW, AT, BE, CH,

CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR,
 BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG
 DE 10129815 A1 20030109 DE 2001-10129815 20010624
 EP 1399551 A2 20040324 EP 2002-750798 20020624
 R: AT, BE, CH, DE, DK, ES, FR, GB, GR, IT, LI, LU, NL, SE, MC, PT,
 IE, SI, LT, LV, FI, RO, MK, CY, AL, TR
 PRAI DE 2001-10129815 A 20010624
 WO 2002-DE2302 W 20020624

AB The invention relates to a method for selectively sepp. bacterial cells
 and/or cell components such as plasmids, whereby the sepp. is carried out
 using a solid support to which bacteriophages or bacteriophage proteins
 are attached. Thus, biotin-labeled phage T4 were immobilized on
 streptavidin-coated magnetic beads and these were used in affinity purifn.
 of E. coli. Plasmids of the purified E. coli may then be isolated.

L5 ANSWER 2 OF 5 BIOTECHDS COPYRIGHT 2004 THE THOMSON CORP. ON STN
 AN 2003-28040 BIOTECHDS
 TI Adapter-directed display system having expression vector with
 sequence encoding exogenous polypeptide and helper vector, for
 displaying exogenous polypeptide e.g., receptor ligand on outer surface
 of genetic package;
 recombinant protein production via plasmid expression in host cell

AU WANG C; ZHONG P; WANG X
 PA WANG C; ZHONG P; WANG X
 PI US 200104355 5 Jun 2003
 AT US 2001-33399 2 Nov 2001
 PRAI US 2001-33399 2 Nov 2001
 DT Patent
 LA English
 OS WPI: 2003-801222 (75)
 DERMENT ABSTRACT:

NOVELTY - An adapter-directed display system for displaying an exogenous
 polypeptide on the outer surface of a genetic package, comprising an
 expression vector with a ***sequence*** that encodes exogenous
 polypeptide fused to first adapter ***sequence***, and a helper
 vector with outer-surface sequences encoding proteins fused to second
 adapter, the polypeptide is produced in a host cell to cause display of
 the polypeptide, is new.

DETAILED DESCRIPTION - An adapter-directed display system (1) for
 displaying an exogenous polypeptide on the outer surface of a genetic
 package, comprising an expression vector comprising a coding
 sequence that encodes the exogenous polypeptide fused in-frame

to a first adapter ***sequence***, where the vector is devoid of
 outer-surface sequences encoding any functional outer-surface proteins of
 the genetic package, and a helper vector comprising outer-surface
 sequences encoding outer-surface proteins necessary for packaging the
 genetic package, where at least one of the outer-surface protein is fused
 in-frame to a second adapter, the first and second adapter acting, when
 the polypeptide is produced in a suitable host cell, to cause the display
 of the polypeptide through pairwise interaction between the first and
 second adapters. INDEPENDENT CLAIMS are also included for the following:

(1) a helper vector (II) for displaying a polypeptide on the outer
 surface of a genetic package comprising, outer-surface sequences
 necessary for packaging the genetic package, where at least one of the
 surface presenting sequences in fused in-frame to an adapter, the adapter
 acting, when the polypeptide is produced in a suitable host cell, to

cause the display of the polypeptide; (2) an expression vector (III) for
 producing a polypeptide within or on the outer surface of a genetic
 package, comprising a coding ***sequence*** encoding the polypeptide
 fused in-frame to a first adapter, where the vector is devoid of
 outer-surface sequences encoding any functional outer-surface proteins of
 the genetic package, and expression of the polypeptide on the outer
 surface of the genetic package is mediated through non-covalent pairwise
 interaction between the first adapter and a second adapter, where the
 second adapter is fused to an outer-surface protein; (3) a kit comprising
 (I), (II) or (III) in suitable packaging; (4) a host cell comprising (I),
 (II) or (III); (5) a polypeptide displayed on the outer surface of a
 genetic package using (I); (6) a genetic package (IV) displaying on its
 outer surface a fusion polypeptide, the fusion polypeptide comprising a
 polypeptide ***sequence*** to be displayed, fused in-frame with a
 first adapter, the first adapter acting, when the fusion polypeptide is
 produced in a suitable host cell, to cause the display of the fusion
 polypeptide through non-covalent pairwise interaction between the first
 adapter and a second adapter that is linked to an outer-surface protein;
 (7) a selectable library comprising several genetic packages, at least
 one being (IV); (8) a selectable library (V) comprising several genetic
 packages, at least one member of several packages displaying a
 polypeptide on its outer surface using (I); and (9) detecting (M1) the
 presence of a specific interaction between a test agent and an exogenous
 polypeptide that is displayed on a genetic package, involves providing a
 genetic package displaying the exogenous polypeptide that is prepared
 using (I), contacting the genetic package with the test agent under
 conditions suitable to produce a stable polypeptide-agent complex, and
 detecting the formation of the stable polypeptide-agent complex on the
 genetic package, thus detecting the presence of a specific interaction.

BIOTECHNOLOGY - Preferred System: (1) is a phage or bacterial
 display system. The genetic package is chosen from viruses, cells and
 spores. The outer-surface sequences encode functional coat proteins of a
 phage. The phage is a filamentous phage. The outer-surface sequences are
 chosen from gene III, gene VI, gene VII, gene VIII and gene IX of a
 filamentous phage. The outer-surface sequences encode bacterial
 outer-surface proteins chosen from Lpp-OmpA, TrpA, Pal, OprI, Imp and
 AIDA-1. The first and second adapters are homodimerization or
 heterodimerization sequences. The homodimerization sequences consist of a
 pair of cysteine residues. The first and second adapters form a
 coiled-coil dimer, where the first and second adapters are leucine
 zippers and comprise heterodimeric receptor sequences that mediate
 heterodimerization of the receptors. The first and second adapters
 comprise heterodimerization sequences of GABAB receptor 1/receptor 2 and
 GABAB receptor 2/receptor 1, respectively. The helper vector further
 comprises at least one additional copy of outer-surface ***sequence***
 that competes for packaging with the fusion outer-surface

sequence. The expression vector is chosen from pBAMX14 and
 pBAMX15. The phage helper vector is chosen from GM-Ultrahelper phage
 vector, GM-Ultrahelper phage vector and GMCT-Ultrahelper phage vector.
 Preferred Vector: (II) is a phage helper vector or bacterial helper
 vector. The adapter causes the display of the polypeptide through
 pairwise interaction with a second adapter, which is fused in-frame with
 the polypeptide. The adapter causes the display of the polypeptide in the
 absence of expression of an outer-surface protein through a phagemid
 vector or a plasmid. The two adapters are heterodimerization or
 homodimerization sequences. (III) is a phagemid vector or bacterial
 expression vector. The outer-surface sequences encode bacterial

outer-surface proteins. Preferred Method: In (M1), the exogenous polypeptide is chosen from antigen-binding unit, cell surface receptor, receptor ligand, cytosolic protein, secreted protein and nuclear protein. Preferably, the exogenous polypeptide is antigen-binding unit. The test agent is chosen from protein, polysaccharides, lipid and their combinations. The test agent is an antigen or ligand.

USE - (1) is useful for displaying a polypeptide on the outer surface of a genetic package, which involves causing (1) to be transcribed and translated in a suitable host cell. (V) is useful for obtaining a polypeptide with desired property, which involves providing (V), and screening the selectable library to obtain at least one genetic package displaying a polypeptide with the desired property. The desired property is binding specificity to an agent of interest. Screening the selectable library further involves isolating the genetic package that displays a polypeptide having the desired property. Isolating the genetic package further involves obtaining a nucleotide. ***sequence*** from the genetic package that encodes the polypeptide with the desired property. The polypeptide with the desired property is chosen from antigen-binding unit, cell surface receptor, receptor ligand, cytosolic protein, secreted protein, nuclear protein, and their functional motif (claimed).

EXAMPLE - Construction of an adapter-directed display system comprising CM-Ultrahelper phage vector was done as follows. The PABM13 vector was constructed by replacing the ***sequence*** between the XbaI and BglII sites of vector PABMD1 with a synthetic DNA fragment comprising 5'-3' gene III leader ***sequence***, a KpnI site, a coding ***sequence*** for Ala-Cys-Gly-Gly and a Myc-tag. The synthetic ***sequence*** was linked in-frame with gene III in PABMD1 vector. The CM-Ultrahelper phage vector was constructed by replacing the KpnI/BamHI fragment encoding and partial p III protein in the KOT Kpn helper vector with the corresponding fragment encoding a partial p III leader and the adapter2-pIII fusion protein from the PABM13 vector. The resulting CM-Ultrahelper phage vector encoded an engineered pIII capsid fused with Cys-Myc domain placed at the N-terminal of pIII. The B9 KOT Kpn helper phage clone was used for the construction of CM-Ultrahelper phage vector. After subcloning of the engineered gene III fragment into KOT Kpn phage vector, 24 kanamycin-resistant colonies were grown in 96-well microtiter plates in 2xYT medium with 70 microg/ml kanamycin, and supernatants were used for phage screening by phage enzyme linked immunosorbent assay (ELISA). Twenty three clones were found to generate phage particles. To confirm the ability of CM-Ultrahelper phages to package the expressed Cys-Myc-pIII fusion into the phage particles, ELISA assay using anti-Myc antibody was performed. The CM-Ultrahelper phages from the selected five clones displayed the Myc-tag on their surfaces. Such Myc-tag was not detectable in the KOT helper phage negative control. The vector PABM15 and was constructed from PABMD2 by replacing the fd gene III fragment of PABMD2 with a synthetic DNA fragment encoding the HA-tag and Gly-Gly-Cys. To demonstrate that a functional protein was displayed using CM-Ultrahelper phage vector, the single-chain antibody AM1 was subcloned into the PABM15 vector. The resulting PABM15-AM1 was transformed into TGI cells, and the cells were superinfected with CM-Ultrahelper phage with a multiplicity of infection (MOI) of 100. Phage particles were generated and purified. The single-chain antibody displayed on the phage surface was detected by phage ELISA using plates coated with AM1-antigen 2x1012 phages were added for each well. The secondary antibody was horse radish peroxidase (HRP)-conjugated anti-M13 antibody. The ELISA results showed that the phagemid particles generated

from all four MOI infections were capable of specifically binding to AM1-antigen, which indicated that functional single-chain antibody was displayed on the phage surface. The control phagemids generated from TGI carrying PABM15-AM1/M13K07 vectors did not bind to AM1-antigen. The phage particles were also used for western blot analysis. Phage particles were denatured by heating in sodium dodecyl sulfate (SDS) sample buffer under non-reducing conditions, the scfv antibody was only displayed by CM-Ultrahelper phage, not by M13K07 helper ***phage***. (****7*** pages)

L5 ANSWER 3 OF 5 DISSABS COPYRIGHT (C) 2004 ProQuest Information and Learning Company; All Rights Reserved on STN
AN 84:22121 DISSABS Order Number: AAR8501756
TI THE ISOLATION AND CHARACTERIZATION OF THE BACILLUS SUBTILIS GENE ENCODING
THE MAJOR SIGMA FACTOR OF RNA POLYMERASE (SIGMA-43, SIGMA-55, RPOD)
AU GITT, MICHAEL ALAN [Ph.D.]
CS UNIVERSITY OF CALIFORNIA, DAVIS (0029)
SO Dissertation Abstracts International, (1984) Vol. 45, No. 11B, p. 3493.
Order No.: AAR8501756. 156 pages.

DT Dissertation
FS DAI
LA English
ED Entered STN: 19921118
AB Last Updated on STN: 19921118

To understand the in vivo role of the major sigma subunit of Bacillus subtilis RNA polymerase, and to understand more about its mechanism of action and regulation, we have isolated its gene, the rpoD gene. Using immunological screening, we isolated 13 phage, all having at least two 3.5 kb EcoRI fragments in common. One representative phage, (lambda)GFW85-sigma-82, contained only the two 3.5 kb inserts in addition to the lambda arms. Its lysate was shown to contain a product which crosses-reacted with anti-sigma-43 (aka sigma-55) antibody, and which was identical to authentic sigma-43 in apparent molecular weight, charge, and VS protease peptide map.

Restriction fragments of the insert DNA were subcloned into plasmid pCP12 to localize the rpoD gene to a 1.7 kb EcoRI-SphI fragment, using a maxicell assay. Other proteins were encoded in the adjacent DNA within the original 7 kb insert. The subcloned 1.7 kb fragment coded for a product which was identical to authentic sigma-43 in apparent molecular weight, peptide map, and immunologic properties.

The 1.7 kb fragment was sequenced and the primary structure of sigma-43 deduced and compared to the N-terminal ***sequence*** of authentic sigma-43. The molecular weight of sigma-43 is calculated as 42,828 daltons, rather than the 55,000 value obtained from gel electrophoresis. I have located a strong ribosomal binding site just upstream of the initiator methionine and a rho-independent transcription terminator ***sequence*** just downstream of the translational termination codon. Codon usage resembles the average for several B. subtilis genes. The rpoD gene appears to be part of an operon, as is the case in E. coli.

The sigma-43 protein exhibits extensive homology, especially in the C-terminal two-thirds of the ***sequence***, to sigma-70 of E. coli. A large internal stretch of 245 amino acids present in sigma-43 is absent in sigma-43. Other proteins exhibiting homology with sigma-43 include the core-binding DNA protein, the heat shock regulator protein of E. coli, and specific RNA-binding proteins. No amino acid homology was found between sigma-43 and B. subtilis phage Sp01 sigma-gp28, E. coli

phage **77*** RNA polymerase, or E. coli DNA primase.

L5 ANSWER 4 OF 5 PROMT COPYRIGHT 2004 Gale Group on STN

AN 2003:538870 PROMT
 TI Maximizing the yield of full-length RNA from an in vitro transcription
 reaction. (Tools & Techniques)
 AU Mele, Ron; Pease, Jim; Kramer, Katharine
 SO Bioscience Technology, (Sept 2003) Vol. 28, No. 9, pp. 8(2).
 PB Reed Business Information
 DT Newsletter
 LA English
 WC 1986

AB *FULL TEXT IS AVAILABLE IN THE ALL FORMAT*
 Abstract
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 90504.

L5 ANSWER 5 OF 5 WPIDS COPYRIGHT 2004 THE THOMSON CORP ON STN
 AN 1987-286263 (41) WPIDS
 CR 1993-257894 (32)
 DMC C1987-121376
 TI Pure nucleic acid comprising G 10 L ***sequence*** - useful for
 enhancing expression of genes in bacteria for prodn. of hormones, plant or
 bacterial enzymes etc..

DC B04 D16
 IN OLINS, P O; OLAFS, P
 PA (MONS) MONSANTO CO
 CYC 21
 PI EP 241446 A 19871014 (198741)* EN 32
 R: AT BE CH DE ES FR GB GR IT LI LU NL SE
 AU 8770661 A 19871001 (198746)
 HU 43625 T 19871130 (198751)
 JP 63000289 A 19880105 (198806)
 DK 8701544 A 19871221 (198810)
 ZA 8702242 A 19871124 (198817)
 EP 241446 B1 19950301 (199513) EN 39
 R: AT BE CH DE ES FR GB GR IT LI LU NL SE
 DE 3751100 G 19950406 (199519)
 ES 2070828 T3 19950616 (199531)
 IL 82017 A 19960119 (199616)
 IE 67890 B 19960501 (199629)
 JP 2511948 B2 19960703 (199631) 32
 CA 1340091 C 19981020 (199901)
 ADT EP 241446 A EP 1987-870039 19870326; JP 63000289 A JP 1987-72974 19870326;
 DE 3751100 G DE 1987-2242 19870326; EP 241446 B1 EP 1987-870039 19870326;
 T3 EP 1987-870039 19870326; IL 82017 A IL 1987-82017 19870326; IE 67890 B
 IE 1987-782 19870326; JP 2511948 B2 JP 1987-72974 19870326; CA 1340091 C
 CA 1987-531052 19870326
 FDT DE 3751100 G Based on EP 241446; ES 2070828 T3 Based on EP 241446; JP
 PRAI 2511948 B2 Previous Publ. JP 63000289 19860327
 AB EP 1987-5821 19870204; US 1986-845159
 EP 241446 A UPAB; 19970202
 (1) Pure nucleic acid molecule comprising a G10L ***sequence*** is

new. The ***sequence*** esp. comprises about the first 100 nucleotides
 immediately 5' to the translation start codon of the ***bacteriophage***
 ****77*** gene 10 coding ***sequence***. (2) Prodn. of a
 heterologous
 protein (1) in bacteria comprises causing expression of a gene comprising
 a G10L ***sequence*** operatively joined to a DNA ***sequence***
 coding for (1).
 USE/ADVANTAGE - The nucleic acid molecule enhances the expression of
 a wide variety of genes, both prokaryotic and eucaryotic, in bacteria. (1)
 is esp. a mammalian growth hormone, atriopeptigen or plant or bacterial
 enzyme.
 Dwg. 0/12

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 BIOTECHNO, CAB, CANCERLIT, CAPLUS, CEABA-VTB, GEN, CIN, CONFSCT, CROPB,
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 2 FILE ESBIOBASE
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 2 QUE (BACTERIOPHAGE OR PHAGE) (W) 77

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 L3
 L4
 L5 22 S L3 NOT PELLETIERER/AU
 5 S L4 AND SEQUENCE

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